

Sympatric Distribution of Three Human *Taenia* Tapeworms Collected between 1935 and 2005 in Korea

Hyeong-Kyu Jeon¹, Kyu-Heon Kim², Jong-Yil Chai³, Hyun-Jong Yang⁴, Han-Jong Rim⁵
and Keeseon S. Eom^{1,*}

¹Department of Parasitology and Medical Research Institute, Chungbuk National University College of Medicine, Cheongju, Chungbuk 361-763, Korea;

²Gyeongin Regional Food and Drug Administration, Incheon 402-835, Korea; ³Department of Parasitology and Tropical Medicine, Seoul National University College of Medicine, and Institute of Endemic Diseases, Seoul National University Medical Research Center, Seoul 110-799, Korea;

⁴Department of Parasitology, College of Medicine, Ewha Womans University, Seoul 158-710, Korea; ⁵Department of Parasitology, Korea University College of Medicine, Seoul 136-705, Korea

Abstract: Taeniasis has been known as one of the prevalent parasitic infections in Korea. Until recently, *Taenia saginata* had long been considered a dominant, and widely distributed species but epidemiological profiles of human *Taenia* species in Korea still remain unclear. In order to better understand distribution patterns of human *Taenia* tapeworms in Korea, partial nucleotide sequences of mitochondrial *cox1* and ITS2 (internal transcribed spacer 2) were determined, along with morphological examinations, on 68 *Taenia* specimens obtained from university museum collections deposited since 1935. Genomic DNA was extracted from formalin-preserved specimens. Phylogenetic relationships among the genotypes (*cox1* haplotype) detected in this study were inferred using the neighbor-joining method as a tree building method. Morphological and genetic analyses identified 3 specimens as *T. solium*, 51 specimens as *T. asiatica*, and 14 specimens as *T. saginata*. Our results indicate that all 3 *Taenia* tapeworms are sympatrically distributed in Korea with *T. asiatica* dominating over *T. saginata* and *T. solium*.

Key words: *Taenia solium*, *T. saginata*, *T. asiatica*, sympatric distribution, molecular identification

INTRODUCTION

The cestodes of medical importance in Korea reportedly include *Taenia* and *Diphyllobothrium* tapeworms. Taeniasis and diphyllobothriasis are among the most prevalent types of human cestode infections in Korea [1]. Human taeniasis has become prevalent in Korea since the first report in 1924 [2]. A nationwide study was reported for the first time in 1969 by Seo et al. [3], and revealed a positive rate of 0.7% for taeniid eggs. Subsequent nationwide surveys conducted every 5 yr by MHSA (Ministry of Health and Social Affairs) and KAHP (Korea Association of Health Promotion) have revealed that the positivity rate for taeniid eggs decreased from 1.9% in 1969 to 0.02% in 1997 and human taeniasis was no more detected in the 2004 KAHP's survey. The infection ratio of *T. solium* versus *T. saginata* was roughly about 1 : 5 based on the collective data from researches reported between 1965 and 1988 [4]. The prevalence of taeniasis among Koreans was considered to be caused by

infection with *T. saginata* until Eom and Rim [5] revised the identification of the Korean *Taenia* tapeworm to establish a new species, *T. asiatica*. Differentiating human-infecting *Taenia* tapeworms is based mostly on the number of uterine branches in the proglottids or on the absence or presence of hooks on the scolex. However, the specificity of differentiation is low for direct observations of these tapeworms due to their morphological similarities, particularly in the eggs and proglottids of adult worms. Differential species identification between *T. saginata* and *T. asiatica* becomes even more difficult when stool sample examination is required because egg morphology is almost indistinguishable under light microscopy.

Epidemiologically, *T. asiatica* has been found in Taiwan, Korea, China, Indonesia, the Philippines, and Vietnam [4,6-11], and the overlapping distributions of the 2 tapeworms in these countries cause frequent confusion. The epidemiological status of the infections with *T. asiatica*, *T. saginata*, and *T. solium* were questionable in Korea, where this has been studied based on raw-food eating habits [4]. Consequently, it was necessary to clarify the distributions of these tapeworms in Korea.

Molecular approaches to differential diagnosis of human

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* Corresponding author (kseom@chungbuk.ac.kr)

Taenia tapeworms have been developed in recent years, including the use of sequence-specific DNA probes, PCR-RFLP (restriction fragment length polymorphisms) [12,13], and multiplex PCR [15,16]. The present study used differences in the morphology, sequence variations in the mitochondrial cytochrome c oxidase I gene, and rDNA internal transcribed spacer II as markers for identifying 3 human *Taenia* tapeworms in Korea.

MATERIALS AND METHODS

Specimens

A total of 68 *Taenia* tapeworms were analyzed in this study (Table 1). These specimens were collected between 1935 and 2005 in the Republic of Korea. Some *Taenia* tapeworm proglottids were passed naturally in stools, or strobilae were discharged after treatment with niclosamide or praziquantel and purgation. The morphological features of the strobilae and proglottids were compatible with those of *Taenia* tapeworms. Mature proglottids were examined by light microscopy with carmine staining on the press-and-fixed specimen in an acetic-acid-formalin-alcohol solution. Twenty-nine specimens were preserved in 10% formalin, and 22 were kept in 70% ethanol, and 17 were kept in -20°C or -70°C.

DNA extraction

Total genomic DNA was extracted from the tissues of terminal proglottids of *Taenia* tapeworms using the DNeasy Tissue Kit (Qiagen, Valencia, California, USA) according to the manufacturer's instructions and used as the template DNA for PCR amplification of the target gene fragments. Genomic DNA of specimens preserved in formalin was extracted using modified Azuma et al., [16] and Li et al., [17] protocols. The tissues (0.01 g) were crushed in liquid nitrogen and then soaked in TE buffer for 3-5 hr and then digested for 30 min in the DNA extraction buffer (100 mM NaCl, 50 mM EDTA in pH 8.0, 50 mM Tris in pH 8.0, 10% SDS in pH 7.2, and 20 mg/ml proteinase K) at 56°C. Samples in a hexadecyltrimethylammoniumbromide (CTAB)/NaCl solution were incubated for 3 hr at 65°C. The cellular debris was removed and the genomic DNA was extracted using an extraction protocol with phenol-chloroform-isoamyl-alcohol (25: 24: 1, v/v/v). DNA was precipitated in 3 M sodium acetate (pH 5.2) and a 2 times volume of 95% ethanol. The pellet was dissolved in 50 µl of TE buffer (10 mM Tris-HCl and 1 mM EDTA in pH 8.0). The genomic DNA was treated with RNase A at 37°C for 30 min and kept at -20°C until use. The

extraction efficiency was examined using agarose-gel electrophoresis. The optical densities at 260 and 280 nm (OD₂₆₀ and OD₂₈₀, respectively) were measured by spectrophotometry.

PCR

The 5.8S-ITS2-28S regions of nuclear ribosome and the mitochondrial *cox1* gene were targeted for PCR amplification. PCR reactions were performed in a reaction mixture of 50 µl with 0.01 µg/µl of genomic DNA, 10 × PCR buffer (20 mM Mg⁺), 10 mM dNTP mixture, 10 pmoles of each primer, and 2.5 U/µl *Taq* DNA polymerase (High Fidelity PCR system, Roche, Mannheim, Germany). PCRs were performed in a GeneAmp PCR System 9700 (Applied Biosystems, Langen, Germany) and involved 1 cycle of initial denaturation at 94°C for 3 min, followed by 35 cycles of denaturation (94°C for 1 min), annealing (46°C for 1 min for *cox1*; 66°C for 30 sec for ITS2), and extension (72°C for 1 min), with a final extension at 72°C for 10 min. The PCR primers used were 687 (5'-GTTAGTTTCTTTCCTC-3') and CS250 (5'-ACCCTGGACGGTGGATCACTCGG-3') for ITS2 which yielded a 534-bp product including 5.8S rDNA and 28S rDNA, corresponding to positions 2723-745 and 3347-3362 of the *Drosophila melanogaster* rDNA sequence (GenBank M21017); and T1F (5'-ATATTTACTTTAGATCATAAGCGG-3') and T1R (5'-ACGAGAAAATATATTAGT CATAAA-3') for *cox1* which yielded a 348-bp product. The PCR amplification products were directly sequenced using DNA sequencer (3730XL, model Applied Biosystems) and the Big-Dye Terminator Cycle Sequencing Reaction Kit (version 3.2, Applied Biosystems).

DNA sequencing and analyses

The sequences of *cox1* and the ITS2 regions from *Taenia* tapeworm specimens were aligned using the CLUSTAL X multiple alignment program [18] and the Bioedit programs (version 5.0.6, BIOSOFT, Ferguson, Missouri, USA). The molecular identification of *Taenia* tapeworm specimens was based on the similarity of nucleotide sequences and phylogenetic relationships with those of *T. asiatica* (GenBank NC 004826), *T. saginata* (GenBank AB066495) and *T. solium* (GenBank NC 004022). Phylogenetic relationships among the *Taenia* genotypes were inferred using different tree-constructing algorithms, i.e., neighbor-joining (NJ), maximum parsimony (MP), and minimum evolution (ME), using the Mega 3.1 program [19]. NJ analysis was performed based on corrected distance matrix of the Kimura's 2 parameter substitution model. Bootstrap analysis was performed with 3,000 replications. The mitochondrial *cox1*

Table 1. Samples analyzed in this study

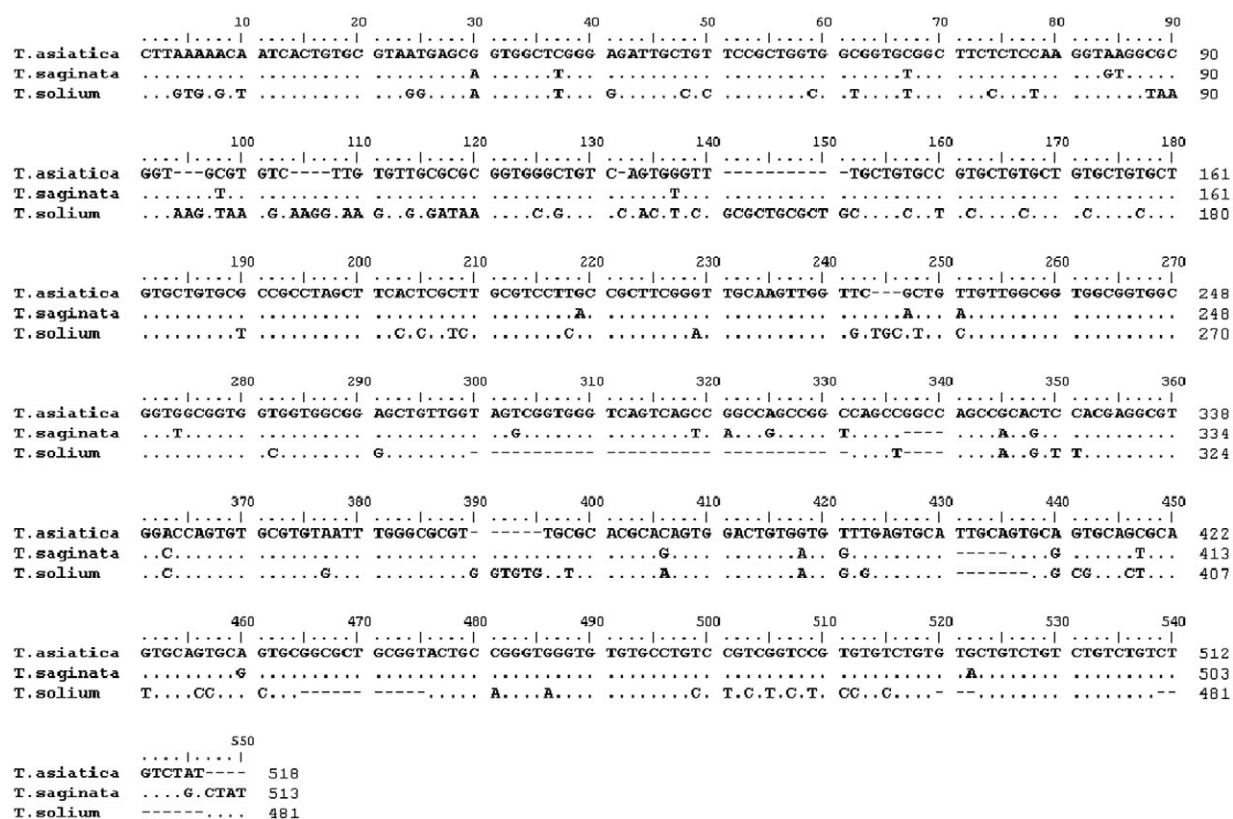
Code	Species	Sex/age	Year	Locality	Fixation condition
G1490	<i>T. asiatica</i>	-	1935	Seoul	10% Formalin
G1485	<i>T. asiatica</i>	M/10	1971	Chungju (Chungbuk)	"
G1496	<i>T. saginata</i>	-	1977	-	"
G1497	<i>T. saginata</i>	M/36	1978	-	"
G1498	<i>T. solium</i>	F/50	1979	Uijeongbu (Gyeonggi)	"
G1494	<i>T. asiatica</i>	F/52	1982	Siheung (Gyeonggi)	"
G1495	<i>T. saginata</i>	M/36	1982	Siheung (Gyeonggi)	"
G1479	<i>T. asiatica</i>	M/48	1982	Yongin (Gyeonggi)	"
G1478	<i>T. saginata</i>	M/56	1983	Nonsan (Chungnam)	"
G1481	<i>T. asiatica</i>	M/51	1983	-	"
G1489	<i>T. asiatica</i>	F/51	1983	Yeoncheon (Gyeonggi)	"
G1482	<i>T. asiatica</i>	M/55	1983	-	"
G1477	<i>T. asiatica</i>	M/46	1984	-	"
G1504	<i>T. saginata</i>	M/45	1984	Pyeongchang (Gangwon)	"
G1507	<i>T. saginata</i>	M/44	1984	Pyeongchang (Gangwon)	"
G1503	<i>T. saginata</i>	M/64	1985	Haenam (Jeonnam)	"
G1487	<i>T. asiatica</i>	-	1986	-	"
G1335	<i>T. asiatica</i>	M/29	1986	Seoul	"
G1506	<i>T. saginata</i>	-	1988	-	"
G1135	<i>T. asiatica</i>	-	1988	Jeju	"
G1336	<i>T. asiatica</i>	-	1989	Cheongju (Chungbuk)	"
K22	<i>T. asiatica</i>	M	1992	Gimcheon (Gyeongbuk)	Frozen
K23	<i>T. asiatica</i>	M/64	1992	Gimcheon (Gyeongbuk)	"
G1483	<i>T. asiatica</i>	F/51	1992	Gyeonggi	10% Formalin
G1101	<i>T. asiatica</i>	-	1993	Chuncheon (Gangwon)	Frozen
G1102	<i>T. asiatica</i>	M	1996	Wanju (Jeonbuk)	"
G1103	<i>T. asiatica</i>	F	1997	Hwasun (Jeonnam)	70% Ethanol
G1104	<i>T. asiatica</i>	M	1997	Hwasun (Jeonnam)	"
G1105	<i>T. asiatica</i>	F	1997	Hwasun (Jeonnam)	"
G1106	<i>T. asiatica</i>	F	1997	Hwasun (Jeonnam)	"
G1107	<i>T. asiatica</i>	F	1997	Hwasun (Jeonnam)	Frozen
G1116	<i>T. asiatica</i>	M	1997	Daegu	70% Ethanol
G1117	<i>T. asiatica</i>	M	1997	Youngju (Gyeongbuk)	"
G1118	<i>T. asiatica</i>	M/57	1997	Wando (Jeonnam)	"
G1119	<i>T. asiatica</i>	F/58	1997	Chuncheon (Gangwon)	70% Ethanol
G1491	<i>T. asiatica</i>	F/57	1997	Wando (Jeonnam)	70% Ethanol
G1138	<i>T. asiatica</i>	M/59	1998	Ansan (Gyeonggi)	"
G1140	<i>T. saginata</i>	F/46	1998	Busan	"
G1484	<i>T. saginata</i>	M/57	2000	-	"
G1190	<i>T. asiatica</i>	M/49	2000	Cheonan (Chungnam)	Frozen
G1192	<i>T. asiatica</i>	-	2000	Jeju	70% Ethanol
G1279	<i>T. asiatica</i>	M/43	2002	Seogwipo (Jeju)	"
G1280	<i>T. asiatica</i>	F/32	2002	Jeju	"
G1292	<i>T. asiatica</i>	M/81	2002	Seogwipo (Jeju)	"
G1299	<i>T. asiatica</i>	F/48	2002	Jeju	"
G1499	<i>T. solium</i>	F/25	-	-	10% Formalin
G1500	<i>T. solium</i>	M/19	-	-	"
G1501	<i>T. saginata</i>	-	-	Yongin (Gyeonggi)	"
K3	<i>T. asiatica</i>	F	-	-	Frozen
K21	<i>T. asiatica</i>	M	-	Jeungpyeong (Chungbuk)	"
K14	<i>T. asiatica</i>	-	-	-	"
K16	<i>T. asiatica</i>	-	-	Seoul	"
K17	<i>T. asiatica</i>	-	-	Seoul	"
K18	<i>T. asiatica</i>	-	-	Seoul	"
K24	<i>T. asiatica</i>	-	-	-	"
K19	<i>T. asiatica</i>	-	-	Seoul	"

(continued next page)

Table 1. (Continued from the previous page) Samples analyzed in this study

Code	Species	Sex/age	Year	Locality	Fixation condition
K36	<i>T. asiatica</i>	-	-	Cheongju (Chungbuk)	"
G1392	<i>T. asiatica</i>	-	-	-	10% Formalin
K41	<i>T. asiatica</i>	-	-	Cheongju (Chungbuk)	Frozen
K39	<i>T. asiatica</i>	-	-	Cheongju (Chungbuk)	10% Formalin
G1486	<i>T. asiatica</i>	M/71	-	-	"
G1488	<i>T. asiatica</i>	-	-	-	"
G1281	<i>T. saginata</i>	-	-	Cheongju (Chungbuk)	Frozen
G1399	<i>T. asiatica</i>	-	2003	Jeju	70% Ethanol
G1400	<i>T. asiatica</i>	-	2003	Jeju	"
G1403	<i>T. saginata</i>	M/37	2003	-	"
G1555	<i>T. asiatica</i>	-	2004	Chuncheon	"
G1579	<i>T. saginata</i>	-	2005	Jeju	"

-, unknown.

Fig. 1. Sequence alignment of ITS2 regions of human *Taenia* tapeworms. Dot (.) indicates identical nucleotide sequence to *T. asiatica*. Gap (-) was introduced for maximum alignments.

gene sequence of *Echinococcus multilocularis* (NC 000928) was used as an outgroup for phylogenetic analysis of *cox1* dataset.

RESULTS

Sequence divergence of *cox1* gene and ITS2

Taenia specimens were molecularly identified by direct sequen-

cing of ITS2 and the partial mitochondrial *cox1* region. ITS2 sequences of human *Taenia* tapeworms were ranged in length from 481 to 512 bp. The various lengths were represented by 503-bp and 512-bp sequences in *T. saginata* and *T. asiatica*, respectively, and 481-bp sequences in *T. solium*. The difference in the number of inferred gap among species ranged between 3 and 36 bp in the human *Taenia* tapeworms (Fig. 1). The nucle-

Table 2. Polymorphic sites of *cox1* sequences of human *Taenia* tapeworms

	115	138	153	174	183	189	190	195	219	225	231	234	237	243	247	252	261	265	267	268	270	273	280	285	288	291	300	315	321	324
Ts	A	G	G	T	G	T	T	A	A	T	A	A	T	C	C	T	T	G	T	T	T	G	T	T	T	C	A	A	A	C
Ta	.	.	T	.	T	C	C	.	G	C	G	.	.	T	T	A	C	T	.	G	G	.
Tsol	T	A	.	A	T	.	C	T	.	.	.	T	A	T	T	C	A	A	A	A	A	A	G	.	.	A	G	.	G	A
	325	330	333	336	338	348	348	355	363	366	381	390	393	399	402	405	408	411	421	422	423	425	426	429	432	438	441	447	450	
Ts	A	T	A	G	C	G	G	T	G	T	G	T	G	G	G	A	A	A	T	A	T	A	C	T	G	C	G	T	G	
Ta	.	.	G	A	T	A	A	G	T	.	.	T	.	C	.	
Tsol	G	G	.	A	T	A	A	G	A	C	G	C	A	A	A	T	G	G	A	G	A	G	T	G	A	T	A	T	T	

Numbers indicate the positions from 5' end of the *cox1* gene, and dots indicate sequence matches with *T. saginata*.
Ts, *T. saginata*; Ta, *T. asiatica*; Tsol, *T. solium*.

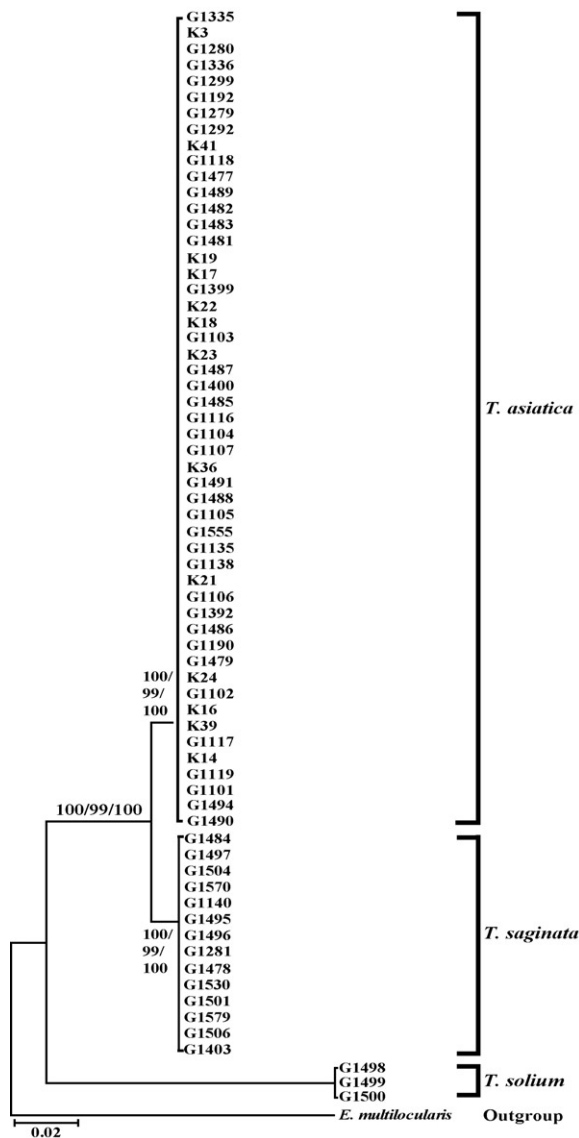


Fig. 2. Phylogenetic trees of human *Taenia* tapeworms based on partial *cox1* sequences inferred from neighbor-joining (NJ), maximum parsimony (MP), and minimum evolution (ME) analyses. Numbers on branches indicate the bootstrap supporting values.

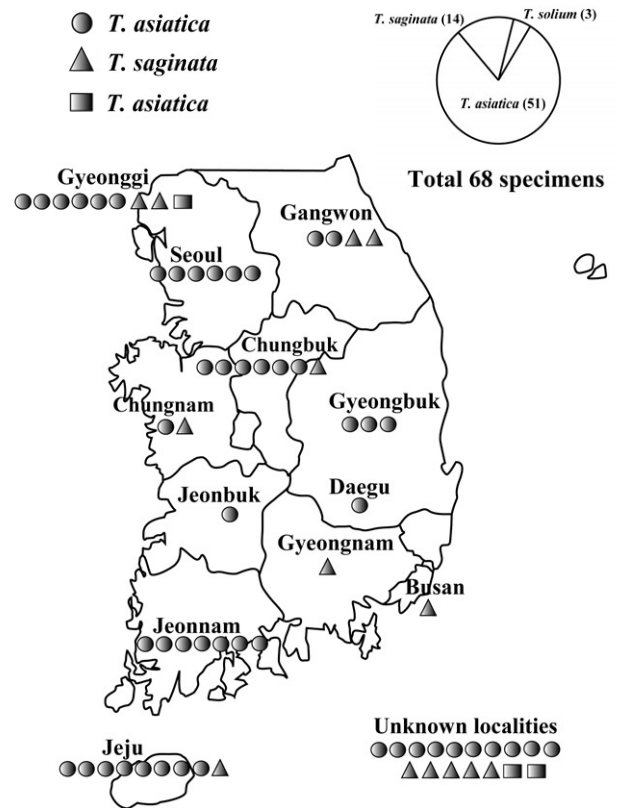


Fig. 3. Species composition and spatial distribution of human infecting *Taenia* tapeworms in Korea.

otide differences in these 3 species varied in size from 12 to 31 bp in the ITS2 region. The sequence divergences of ITS2 between 3 *Taenia* species ranged from 7.3% (*T. saginata* and *T. asiatica*) and 33.7% (*T. asiatica* and *T. solium*), respectively. The *cox1* sequences (348 bp) showed 59 polymorphic sites, with 28 sites of nonsynonymous substitutions in human *Taenia* tapeworms (Table 2). The sequences of *T. asiatica* and *T. saginata* differed by 7%, while those of *T. saginata* and *T. solium* differed by 15%. Phylogenetic analysis of the mtDNA *cox1* sequences for a total

of 68 isolates (of which 51 were obtained from 9 identified provinces in Korea) identified *T. solium* as basal to the *T. saginata*-*T. asiatica* clade. Trees topology using various analytic methods (NJ, MP, and ME) generated was identical with very high confidence values (bootstrap values of 100%, 99%, and 100% in NJ, MP, and ME, respectively) for 3 major branches representing each of 3 species (Fig. 2).

Species composition and spatial distribution

Morphological and genetic analyses identified 3 specimens (from Gyeonggi and unknown localities) as *Taenia solium*, 51 specimens (from Chungbuk, Gangwon, Gyeonggi, Gyeongbuk, Jeonnam, Jeonbuk, Chungnam, Jeju, Seoul, Daegu, and unknown localities) as *T. asiatica* and 14 specimens (from Chungbuk, Gangwon, Gyeonggi, Chungnam, Busan, and unknown localities) as *T. saginata*. Most provinces exhibited 1 or 2 kinds of tapeworms except for Gyeonggi province that showed all 3 species of tapeworms. Ratios of *T. solium*: *T. asiatica*: *T. saginata* calculated from both morphological and molecular data was 3: 51: 14 (or approximately 1: 17: 5). *T. solium*, *T. saginata*, and *T. asiatica* tapeworm infections were only exhibited in Gyeonggi province. *T. saginata* and *T. asiatica* tapeworm infections were presented in Gangwon, Chungbuk, Chungnam and Jeju provinces (Fig. 3). This result indicates that *T. asiatica* was the dominant human-infecting *Taenia* tapeworm in Korea.

DISCUSSION

Two types of taenias (*T. saginata* and *T. solium*) were only recognized in Korea until 1992, with *T. asiatica* being newly described in 1993 as the third type of causative agent. *T. saginata* is morphologically distinct from *T. asiatica* by unarmed restellum, large number of uterine buds, and a posterior protuberance on the proglottids. Nevertheless, differential diagnosis of these species is often a task of specialists with much experience, especially when this is performed in field research conditions, which is hampered by the shortage of parasite materials obtained for examination or variable fixation/preservation conditions of the adult worms collected. These 2 species also exhibit different life cycles: with *T. asiatica* uses suids as intermediate hosts, whereas *T. saginata* utilizes bovids as intermediate hosts in order to complete their life cycles. This criterion, however, is not applicable in the laboratory condition. Even though the epidemiological understanding of *T. asiatica* infection remained questionable, *T. asiatica* rather than *T. saginata*, had been con-

sidered to be the predominant species in Korea [4]. It was therefore necessary to quantify the prevalence of this tapeworm as well as those of *T. saginata* and *T. solium*. There was no *Taenia* egg positive case in the last nationwide survey of the presence of helminth eggs among Koreans, implemented in 2004, suggesting that the occurrence of *Taenia* species is rapidly decreasing. The difficulties of finding out *Taenia*-tapeworm-infected individuals prompted us to plan to analyze collections available over all time, dating back up to 1935 from laboratories and institutes, and including any sources of tapeworms in Korea. Recent advancements in DNA analysis techniques made it possible to differentiate species precisely, but the results were not consistent with epidemiological finding.

Twenty-nine of the 68 examined specimens were preserved in 10% formalin, which can cause cross-linking between DNA and protein, and also degrade DNA by reducing sugar, and inflicting oxidative and hydrolytic damage. Therefore, the DNA recovery yield is usually lower for formalin-fixed specimens than ethanol-fixed or frozen specimen. In this study, genomic DNA was extracted from the modified protocol of Azuma et al. [16] and Li et al. [17]. We extended the time of incubation up to 24 hr, and SDS and proteinase K were added during the incubation process. Most of the formalin-preserved samples yielded weak or no PCR amplification, but secondary PCR using the PCR products obtained from the first round of PCR amplifications as a template make it possible to use the PCR product for direct sequencing. The longest time period of formalin preservation used for molecular identification was the specimen (labeled as *T. saginata* by the original collector) that had been isolated, and kept in 10% formalin since 1935 in the Department of Parasitology, Seoul National University. Very interestingly, *cox1* sequence analysis of the specimen identified it as *T. asiatica*. Overall results were 3 *T. solium*, 51 *T. asiatica*, and 14 *T. saginata*. These results clearly indicate that all 3 human *Taenia* tapeworms are sympatrically distributed in Korea, with *T. asiatica* infections accounting for three-quarters of the total *Taenia* infections.

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REFERENCES

1. Min DY. Cestode infections in Korea. Korean J Parasitol 1990; 28(suppl): 123-144.
2. Inoba J. Human taeniasis caused by eating meat in Chosen. Chuo Juikai Zasshi 1924; 37: 229-239.
3. Seo BS, Rim HJ, Loh IK. Study on the status of helminthic infections in Koreans. Korean J Parasitol 1969; 7: 53-70.
4. Eom KS, Rim HJ. Epidemiological understanding of *Taenia* tapeworm infections with special reference to *Taenia asiatica* in Korea. Korean J Parasitol 2001 39: 267-283.
5. Eom KS, Rim HJ. Morphologic descriptions of *Taenia asiatica* sp. n. Korean J Parasitol 1993; 31: 1-6.
6. Eom KS, Jeon HK, Kong Y, Hwang UW, Yang Y, Li X, Xu L, Feng Z, Pawlowski ZS, Rim HJ. Identification of *Taenia asiatica* in China: molecular, morphological and epidemiological analysis of a Luzhai isolate. J Parasitol 2002; 88: 758-764.
7. Ito A, Nakao M, Wandra T. Human taeniasis and cysticercosis in Asia. Lancet 2003; 362: 1918-1920.
8. Anantaphruti MT, Yamasaki H, Nakao M, Waikagul J, Watthanakulpanich D, Nuamtangong S, Maipanich S, Pubampen S, Sanguankiat S, Muennoo C, Nakaya K, Sato MO, Sako Y, Okamoto M, Ito A. Sympatric occurrence of *Taenia solium*, *T. saginata*, and *T. asiatica*, Thailand. Emerg Infect Dis 2007; 13: 1413-1416.
9. Fan PC, Lin CY, Chen CC, Chung WC. Morphological description of *Taenia saginata asiatica* (Cyclophyllidae: Taeniidae) from man in Asia. J Helminthol 1995; 69: 299-303.
10. Simanjuntak GM, Margono SS, Okamoto M, Ito A. Taeniasis/cysticercosis in Indonesia as an emerging disease. Parasitol Today 1997; 13: 321-323.
11. Willingham AL 3rd, De NV, Doanh NQ, Cong Le D, Dung TV, Dorny P, Cam PD, Dalsgaard A. Current status of cysticercosis in Vietnam. Southeast Asian J Trop Med and Public Health 2003; 34(suppl 1): 35-50.
12. Bowles J, McManus DP. Genetic characterization of the Asian *Taenia*, a newly described taeniid cestodes of human. Am J Trop Med and Hyg 1994; 50: 33-44.
13. Yamasaki H, Nakao M, Sako Y, Nakaya K, Sato MO, Mamuti W, Okamoto M, Ito A. DNA differential diagnosis of human taeniid cestodes by base excision sequence scanning thymine-base reader analysis with mitochondrial genes. J Clin Microbiol 2002; 40: 3818-3821.
14. Gonzales LM, Montero E, Morakote N, Pueute S, Diaz De Tuesta JL, Serra T, Lopez-Velez R, McManus DP, Harrison IJS, Parkhouse RME, Garate T. Differential diagnosis of *Taenia saginata* and *Taenia saginata asiatica* taeniasis through PCR. Diagn Microbiol and Infect Disease 2004; 49: 183-188.
15. Yamasaki H, Allan JC, Sato MO, Nakao M, Sako Y, Nakaya K, Qiu D, Mamuti W, Craig PS, Ito A. DNA differential diagnosis of taeniasis and cysticercosis by multiplex PCR. J Clin Microbiol 2004; 42: 548-553.
16. Azuma H, Okamoto M, Oku Y, Kamiya M. Intraspecific variation of *Taenia taeniaeformis* as identified by various criteria. Parasitol Res 1995; 81: 103-108.
17. Li J, Liao X, Yang H. Molecular characterization of a parasitic tapeworm (*Ligula*) based on DNA sequences from formalin-fixed specimens. Biochem Genet 2000; 38: 309-322.
18. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG. The CLUSTAL X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. Nucleic Acids Res 1997; 25: 4876-4882.
19. Kumar S, Tamura K, Nei M. MEGA3: Integrated software for molecular evolutionary genetics analysis and sequence alignment. Brief Bioinformatics 2004; 5: 150-163.

