

Fish-borne Zoonotic Trematode Metacercariae in the Republic of Korea

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Abstract: The prevalence of fish-borne trematodes (FBT), including *Clonorchis sinensis*, is still high in riverside areas of the Republic of Korea. The author reviewed the detection and identification methods, differential keys, fish intermediate hosts, and morphological characteristics of FBT metacercariae. FBT metacercariae found in freshwater fish are classified mainly into 4 families, i.e., Opisthorchiidae, Heterophyidae, Echinostomatidae, and Clinostomidae. The metacercariae of *C. sinensis*, found in 40 species of freshwater fish, are elliptical and $0.15\text{--}0.17 \times 0.13\text{--}0.15$ mm in size, have nearly equal sized oral and ventral suckers, brownish pigment granules, and an O-shaped excretory bladder. Their general morphologies are similar to those of *Metorchis orientalis* (except in the thickness of the cyst wall). *Metagonimus* spp. (*M. yokogawai*, *M. takahashii*, and *M. miyatai*) metacercariae are subglobular or disc-shaped, and 0.14–0.16 mm in diameter. They have yellow-brownish pigment granules, a ventral sucker defectively located from median, and a V-shaped excretory bladder. The metacercariae and fish intermediate hosts of *Centrocestus armatus*, *Clinostomum complanatum*, and 3 echinostomatid flukes (*Echinostoma hortense*, *E. cinetorchis*, and *Echinochasmus japonicus*) were summarized. FBT metacercariae detected in brackish water fish are mainly members of the Heterophyidae. The morphological characters, identification keys, and fish intermediate hosts of 7 species (*Heterophyes nocens*, *Heterophyopsis continua*, *Pygidiopsis summa*, *Stellantchasmus falcatus*, *Stictodora fuscata*, *Stictodora lari*, and *Acanthotrema felis*) were also reviewed. The contents treated in this study will provide assistance at the laboratory bench level to those working on recovery of metacercariae from fish hosts and identifying them.

Key words: fish-borne trematode (FBT), metacercaria, freshwater fish, brackish water fish, identification, differential key

INTRODUCTION

The fish-borne trematode (FBT) infections affect the health of more than 18 million people around the world, particularly in Asian countries. Humans are mainly infected with FBT when they eat raw or inadequately cooked fish contained infective larvae, metacercariae. These flukes provoke remarkable morbidity and cause serious damage to aquaculture, which is a valuable source of food and employment in developing countries. Freshwater and brackish water fish play a major role as the source of human infections with foodborne trematodes, which are receiving increasing attention as the information on their diversity and prevalence emerges in some Asian countries [1-3]. Whereas, some other kinds of animals, i.e., marine fish, mollusca, crustaceae, insects, amphibia and reptiles, act less commonly as the infection source or second intermediate hosts of trematodes. Accordingly, elimination of these parasites from the food supply, especially fish, is a very important work in epidemiological points

of view.

The liver flukes, *Clonorchis sinensis* and *Opisthorchis* spp. (not existing in Korea), have been known as the representative FBT. In addition, many species of intestinal flukes, mainly those belonging to the Heterophyidae and Echinostomatidae are also contracted to humans by eating raw fish. In the Republic of Korea, many species of FBT including *C. sinensis* have been reported. *C. sinensis* is still prevalent in riverside areas and is the most important helminth species in aspects of public health. About 11 species of the Heterophyidae and 3 species of the Echinostomatidae have been reported as the intestinal flukes infected by eating raw fish meat [2-5].

Recent trends of helminthic infection in the Republic of Korea are characterized by a remarkable decrease of soil-transmitted nematodiasis and moderate endemicity of foodborne trematodiasis. Especially, the prevalence of FBT infections, such as *C. sinensis* and intestinal flukes, has maintained at relatively high levels in riverside areas [5-7]. Therefore, it is worthwhile to pay attention to the infective stage and source of human infections with FBT in epidemiological points of view. In this paper, the FBT metacercariae of medical importance in the Republic of Korea are briefly reviewed to provide assistance at the laborato-

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ry bench level to those working for recovery of metacercariae from the second intermediate hosts.

DETECTION OF METACERCARIAE

Examination of FBT metacercariae in the second intermediate hosts is commonly done by 2 methods, i.e., muscle compression and pepsin-HCl artificial digestion techniques. The compression method is done by following step-by-step procedures.

1. Identify the species of fish.
2. Weigh and measure the size of each fish and record.
3. Take some of the flesh from different parts of fish (e.g., head, gill, muscles, fin, scale, intestine, other viscera).
4. Weigh each sample to estimate metacercarial density.
5. Compress each sample between 2 glass slides (repeat 2-3 times to increase the detection rate).
6. Observe and identify metacercariae under stereomicroscopy.
7. Count the number of metacercariae and calculate density.

On the other hand, the artificial digestion method is more complicated, and its step-by-step procedures are as follows.

1. Identify the species of fish.
2. Weigh and measure the size of each fish and record.
3. Take some of the flesh from different parts of fish (e.g., head, gill, muscles, fin, scale, viscera). Whole fish can be examined at one time.
4. Grind fish flesh one by one (or small fish can be grouped to be ground at one time) in a mortar with pestle.
5. Transfer the ground sample into a beaker containing artificial gastric juice (conc. HCl 8 ml + pepsin 1:10,000 6 g + distilled water 1,000 ml).
6. Mix well and place in a 37°C incubator for 2 hr (or longer for hard parts like fin or scale) with occasional stirring.
7. Remove the larger particles (bones, scales, fins, and undigested materials) by the filtration (1 × 1 mm of mesh) of digested materials.
8. Add 0.85% saline, and let it stand for a while.
9. Discard supernatant very carefully and keep the sediment.
10. Repeat procedures 8 and 9 several times until the supernatant becomes clear.
11. Transfer the sediment a small bit into a Petri dish containing 6-7 ml physiological saline.
12. Observe and identify metacercariae using a stereomicroscope and light microscope (see Identification Method).
13. Isolate the metacercariae and put into a small dish.
14. Count the number of metacercariae of each fluke species.

15. Prepare for experimental infection to laboratory animals.

16. Store in a refrigerator until use.

Advantages of the compression method include being able to know the exact location and infection site of metacercariae in fish examined and economical without use of expensive reagents, such as pepsin. Whereas, in the digestion method, a large amount of samples can be dealt with, metacercariae can be isolated and purified, excellent morphologies can be obtained and more easily identified, and exact numbers of metacercariae can be prepared for experimental infections.

IDENTIFICATION OF METACERCARIAE

For the identification of FBT metacercariae, the following general procedures will be help. First of all, collect separately similar-shaped metacercariae based on the general feature in a small Petri dish. Second, move them with a spoid on a glass slide, cover a cover slip, and observe detailed morphology under a light microscope. Finally, identify the metacercariae based on characteristic features and dimensions. As the characteristic features, the shape of cysts, presence and size of suckers, and shape and contents of excretory bladder are important.

On the other hand, if the morphological features of metacercariae are not obvious and difficult to see, it should be excysted using the techniques described below. When the cyst wall is very thin, metacercariae can easily be liberated by giving only a slight pressure on the cover slip. Otherwise (when the cyst wall is thick and elastic), the artificial digestion methods are recommended. There are several published procedures for digesting the cyst and freeing the metacercariae. Basically, they involve incubating the metacercariae in trypsin or bile at 37°C for a brief interval [8-12].

Metacercariae in freshwater fish in the Republic of Korea

More than 16 species of digenetic trematode metacercariae in 7 families have been found in freshwater fish from the Republic of Korea (Table 1). Among them, 8 species in 4 families, Opisthorchiidae (*Clonorchis sinensis*), Heterophyidae (3 *Metagonimus* spp. and *Centrocestus armatus*), Echinostomatidae (*Echinostoma hortense*, *E. cinetorchis*, and *Echinochasmus japonicus*) and Clinostomidae (*Clinostomum complanatum*), are infected by eating raw flesh of freshwater fish in the Republic of Korea.

Morphological characters of metacercariae in freshwater fish

The metacercariae of *C. sinensis* are elliptical and 0.15-0.17 × 0.13-0.15 mm in size, have nearly equal sized oral and ventral

suckers, brownish pigment granules, and an O-shaped excretory bladder. Their general morphologies are similar to those of *Opisthorchis viverrini*, which is prevalent in Thailand, Vietnam, and Lao PDR, except the size of cysts ($0.19\text{--}0.25 \times 0.15\text{--}0.22$ mm) (Fig. 1) and to those of *Metorchis orientalis* ($0.156\text{--}0.188 \times 0.138\text{--}0.170$ mm in size) except the thickness of the cyst wall ($5\text{--}19\text{ }\mu\text{m}$) (Fig. 2). The metacercariae of *C. sinensis* can be differentiated from those of *Metagonimus* spp., although the size and shape of cysts are similar each other; the size and location of suckers are somewhat different (Fig. 3).

A large number of *Metagonimus* sp. metacercariae are encysted in the fin and scale of fish intermediate hosts. They are subglobular or disc-shaped and $0.14\text{--}0.16$ mm in diameter, and have yellow-brownish pigment granules, a ventral sucker defectively located from median, and a V-shaped excretory bladder. The

Table 1. Zoonotic metacercariae encysted in freshwater fish in the Republic of Korea

Family	Species
Opisthorchiidae	<i>Clonorchis sinensis</i> , <i>Metorchis orientalis</i> , <i>M. taiwanensis</i>
Heterophyidae	<i>Metagonimus yokogawai</i> , <i>M. miyatai</i> , <i>M. takahashii</i> , <i>Centrocestus armatus</i>
Echinostomatidae	<i>Echinostoma hortense</i> , <i>E. cinetorchis</i> , <i>Echinochasmus japonicus</i>
Clinostomidae	<i>Clinostomum complanatum</i>
Cyathocotylidae	<i>Cyathocotyle orientalis</i> , <i>Holostephanus nipponicus</i>
Cryptogonimidae	<i>Exorchis oviformis</i> , <i>Pseudexorchis major</i>
Bucephalidae	<i>Dollfustrema echinatum</i>

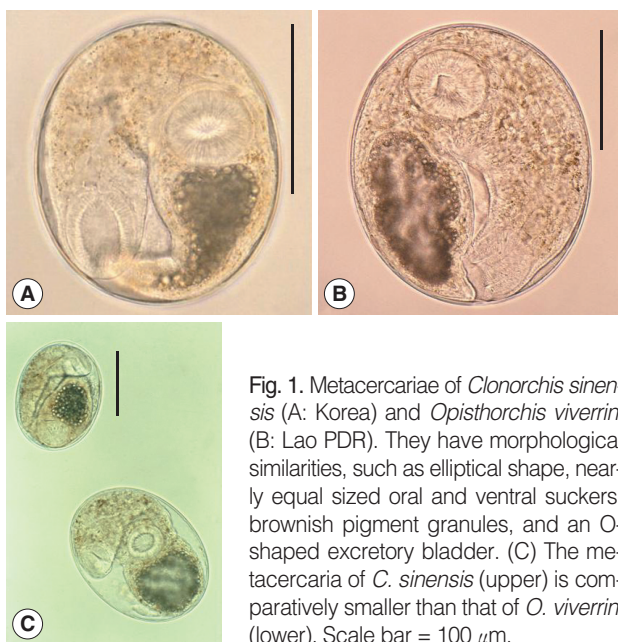


Fig. 1. Metacercariae of *Clonorchis sinensis* (A: Korea) and *Opisthorchis viverrini* (B: Lao PDR). They have morphological similarities, such as elliptical shape, nearly equal sized oral and ventral suckers, brownish pigment granules, and an O-shaped excretory bladder. (C) The metacercaria of *C. sinensis* (upper) is comparatively smaller than that of *O. viverrini* (lower). Scale bar = $100\text{ }\mu\text{m}$.

metacercariae of *Metagonimus* spp. cannot be differentiated morphologically at a species level. Due to host specificity, *M. yokogawai* metacercariae are dominantly collected from sweetfish, *Plecoglossus altivelis*, *M. miyatai* metacercariae are detected in the pale chub, *Zacco platypus* and dark chub, *Z. terminckii*, and *M. takahashii* metacercariae are found in the crucian carp, *Carassius auratus* (Fig. 4). The metacercariae of another heterophyid fluke, *Centrocestus armatus*, are frequently encysted in the viscera of fish

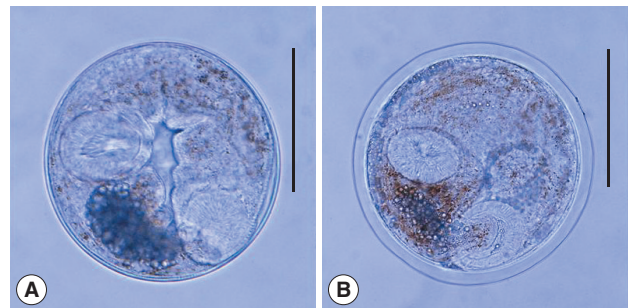


Fig. 2. Metacercariae of *Clonorchis sinensis* (A) and *Metorchis orientalis* (B). (C) They are morphologically very similar except the thickness of the cyst wall. *M. orientalis* metacercaria (lower) is globular, $0.16\text{--}0.18$ mm in diameter with a double layered cyst wall (outer cyst wall being 4 times as thick as that of *C. sinensis*; upper), and have nearly equal sized oral and ventral suckers and an O-shaped excretory bladder. Scale bar = $100\text{ }\mu\text{m}$.

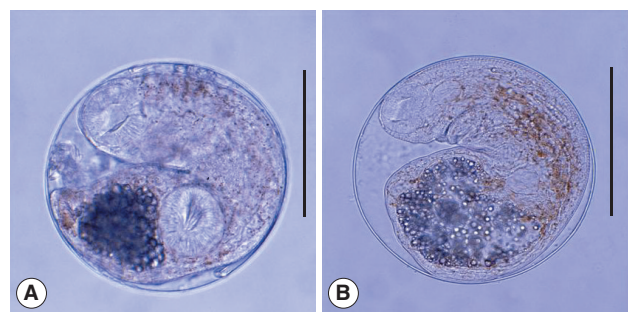
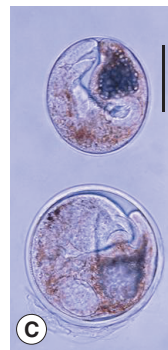
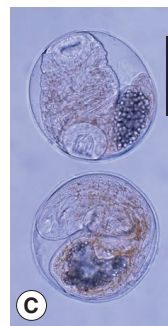


Fig. 3. Metacercariae of *Clonorchis sinensis* (A) and *Metagonimus* sp. (B). (C) They are similar in size, and have yellow-brownish pigment granules in the body. However, the size of the ventral sucker and excretory bladder are somewhat different. Scale bar = $100\text{ }\mu\text{m}$.



hosts. They are elongated and elliptical, $0.20\text{--}0.25 \times 0.10\text{--}0.12$ mm in size, have 40–44 circumoral spines arranged in 2 rows around the oral sucker, and X-shaped excretory bladder (Fig. 5).

Three species of echinostome metacercariae, i.e. *Echinostoma*

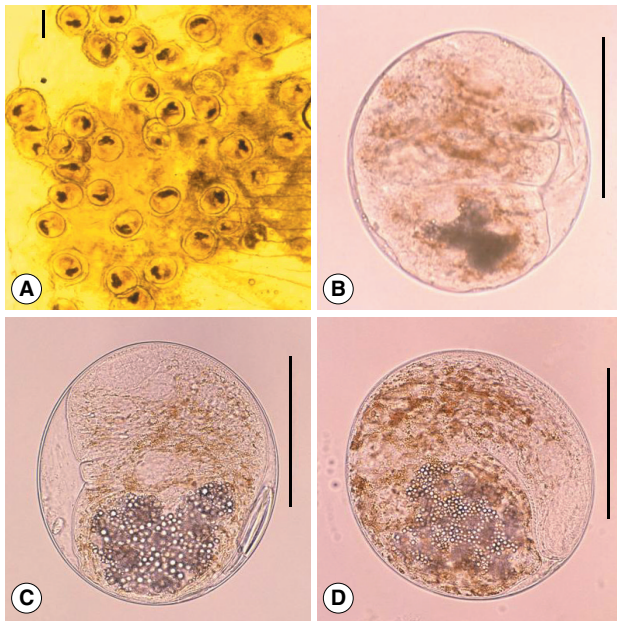


Fig. 4. *Metagonimus* spp. metacercariae. (A) Numerous metacercariae of *M. yokogawai* encysted on a scale of sweetfish, *Plecoglossus altivelis*. Scale bar = $200\ \mu\text{m}$. (B) A *M. yokogawai* metacercaria detected from a sweetfish. Scale bar = $100\ \mu\text{m}$. (C) A *M. miyatai* metacercaria collected from a pale chub, *Zacco platypus*. Scale bar = $100\ \mu\text{m}$. (D) A *M. takahashii* metacercaria detected from a crucian carp, *Carassius auratus*. They are subglobular or disc-shaped, and have yellow brownish pigment granules, a ventral sucker defectively located from median and a V-shaped excretory bladder. Scale bar = $100\ \mu\text{m}$.

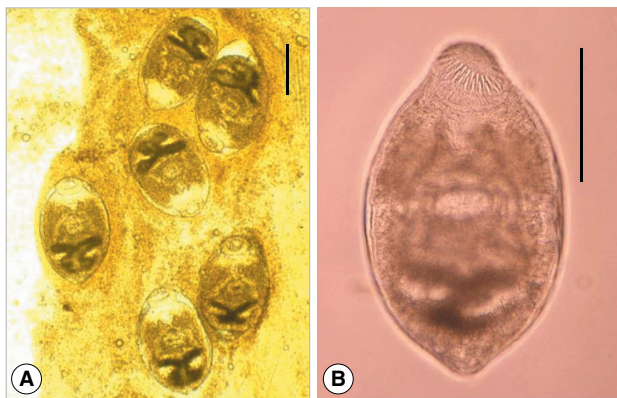


Fig. 5. (A) *Centrocestus armatus* metacercariae encysted in the viscera of the pale chub, *Z. platypus*. (B) They are elongated elliptical and have 40–44 circumoral spines around the oral sucker arranged in 2 rows and an X-shaped excretory bladder. Scale bar = $100\ \mu\text{m}$.

hortense, *E. cinetorchis*, and *Echinochasmus japonicus*, are found in freshwater fish. They are characterized by an oral sucker surrounded by prominent collar spines and excretory granules in 2 rows of excretory tubes. *E. hortense* metacercariae are globular or elliptical, $0.17\text{--}0.19 \times 0.15\text{--}0.16$ mm in size, and have a double layered cyst wall, 27–28 collar spines, and a ventral sucker transversely elliptical, 2-fold as larger as the oral sucker (Fig. 6). *E. cinetorchis* metacercariae are rounded, $0.13\text{--}0.14$ mm in diameter, and have 37 collar spines around the oral sucker and a ventral sucker lying in the middle of the body and larger than the oral sucker (Fig. 7). *E. japonicus* metacercariae are mainly encysted in the gill filaments of fish intermediate hosts. They are elliptical and very small, $0.077\text{--}0.080 \times 0.054\text{--}0.060$ mm in size, have a transparent and double layered cyst wall, 24 dorsally interrupted collar spines, and a ventral sucker lying median at pos-

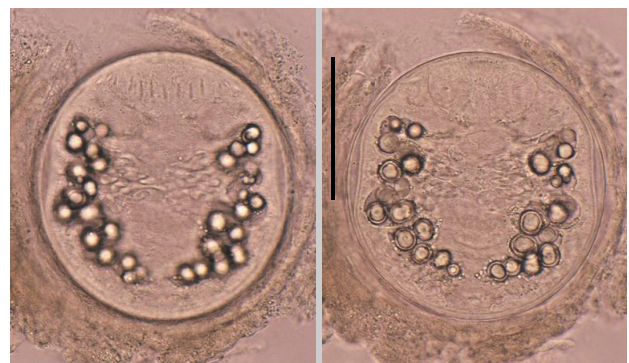


Fig. 6. (A) *Echinostoma hortense* metacercaria detected from a yellowfin goby, *Acanthogobius flavimanus*. It is globular or elliptical in shape, have a double layered cyst wall, 27–28 collar spines and a ventral sucker transversely elliptical, 2-fold as larger as the oral sucker. Scale bar = $100\ \mu\text{m}$.

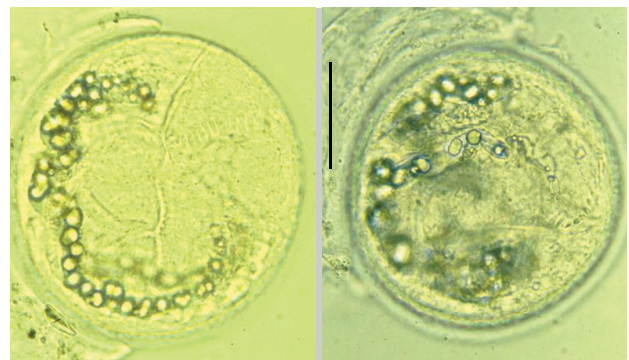


Fig. 7. *Echinostoma cinetorchis* metacercariae collected from a muddy loach, *Misgurnus anguillicaudatus*. They are rounded and have 37 collar spines around the oral sucker and a ventral sucker lying in the middle of the body and being larger than the oral sucker. Scale bar = $50\ \mu\text{m}$.

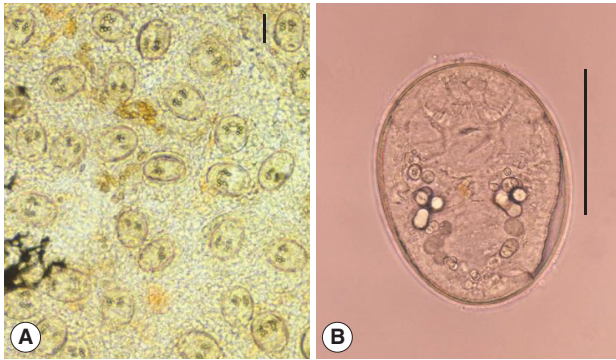


Fig. 8. (A) *Echinochasmus japonicus* metacercariae encysted on the gills of *Pseudorasbora parva*. (B) They are elliptical and very small, have a transparent and double layered cyst wall, 24 dorsally interrupted collar spines, and a ventral sucker lying median at posterior 1/3 of the body. Scale bars = 50 μ m.

terior 1/3 of the body and somewhat smaller than the oral sucker (Fig. 8).

The metacercariae of *C. complanatum* have a very thin cyst wall, and thus they are easily liberated from the cysts during the process of collection. The excysted metacercariae are big (3.28-4.27 \times 0.94-1.46 mm), tongue-shaped, and progenetic (Fig. 9).

Differential keys for metacercariae in freshwater fish

- 1.a. Brownish pigment granules present in 2
- 1.b. Brownish pigment granules absent in body 5
- 2.a. Cyst wall thin or thick; ventral sucker prominent, approximately same size as oral sucker 3
- 2.b. Cyst wall relatively thin; ventral sucker defectively located from median *Metagonimus* spp.
- 3.a. Cyst wall relatively thin *Clonorchis sinensis*
- 3.b. Cyst wall very thick 4
- 4.a. Outer cyst wall below 30 μ m thickness
..... *Metorchis orientalis*
- 4.b. Outer cyst wall over 50 μ m thickness *M. taiwanensis*
- 5.a. Spines present around the oral sucker 6
- 5.b. Spines absent around the oral sucker 9
- 6.a. Spines arranged in 2 rows just around the oral sucker, excretory bladder x-shaped *Centrocestus armatus*
- 6.b. Spines arranged in a single or 2 row, granules in excretory bladder arranged in 2 rows of tube 7
- 7.a. Arrangement of collar spines dorsally interrupted
..... *Echinochasmus* spp.
- 7.b. Arrangement of collar spines not interrupted 8
- 8.a. No. of collar spines: 27 *Echinostoma hortense*
- 8.b. No. of collar spines: 37 *Echinostoma cinetorchis*

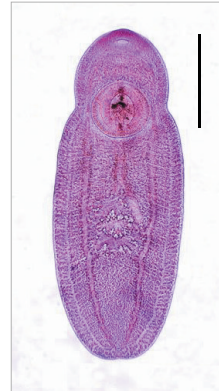


Fig. 9. Excysted metacercaria of *Clinostomum complanatum* found from a short barbel gudgeon, *Squalidus japonicus coreanus* (Semichon's acetocarmine stained), and is big and progenetic. It has a very thin cyst wall, and thus it is easily liberated from the cyst during the process of collection. Scale bar = 100 μ m.

Table 2. Zoonotic metacercariae encysted in brackish water fish in the Republic of Korea

Family	Species
Heterophyidae	<i>Heterophyes nocens</i> , <i>Heterophyopsis continua</i> , <i>Pygidiopsis summa</i> , <i>Sellantchasmus falcatus</i> , <i>Metagonimus takahashii</i> , <i>Stictodora fuscata</i> , <i>Stictodora lari</i> , <i>Acanthotrema felis</i>
Echinostomatidae	<i>Echinostoma hortense</i>
Bucephalidae	<i>Prosorhynchus uniporus</i>

- 9.a. Excretory bladder distributed in the whole body 10
- 9.b. Excretory bladder mainly located in the posterior body 11
- 10.a. Rounded, with thick cyst wall and w-shaped excretory bladder *Cyathocotyle orientalis*
- 10.b. Elliptical, with thin cyst wall and 8-shaped excretory bladder *Holostephanus nipponicus*
- 11.a. Excretory bladder occupying large part of posterior body, and V-shaped *Matacercaria hasegawai*
- 11.b. Excretory bladder Y or V-shaped, and it's arms extended to pharynx-level 12
- 12.a. Excretory bladder V-shaped, with a pair of eye spots
..... *Exorchis oviformis*
- 12.b. Excretory bladder Y-shaped *Pseudexorchis major*

Metacercariae in brackish water fish in the Republic of Korea

More than 10 species of digenetic trematode metacercariae in 3 families have been found in brackish water fishes in the Republic of Korea (Table 2). Among them, 9 species in 2 families, Heterophyidae (*Heterophyes nocens*, *Heterophyopsis continua*, *Pygidiopsis summa*, *Sellantchasmus falcatus*, *Metagonimus takahashii*, *Stictodora fuscata*, *Stictodora lari*, and *Acanthotrema felis*), and Echinostomatidae (*Echinostoma hortense*), are listed as FBT in the Republic of Korea.

Morphological characters of metacercariae in brackish water fish

The metacercariae of *H. nocens* are round or elliptical, 0.13-0.22 × 0.08-0.17 mm in size, with brownish pigment scattered throughout body, a ventral sucker elliptical and larger than the oral sucker, a genital sucker elliptical and lying closely right lateral posteriorly to the ventral sucker, and an excretory bladder O-shaped (Fig. 10A). *H. continua* metacercariae are rounded, 0.38 mm in diameter, and have a cyst wall of about 6 µm thick, a ventral sucker larger than the oral sucker, a genital sucker elliptical and lying dextroposteriorly to the ventral sucker, and an excretory bladder Y-shaped (Fig. 10B). *P. summa* metacercariae

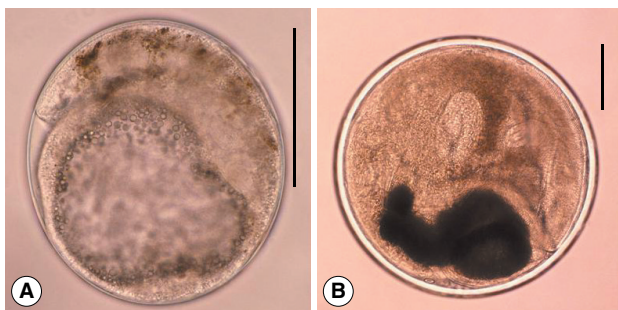


Fig. 10. (A) *Heterophyes nocens* metacercaria detected from a yellowfin goby, *A. flavimanus*. It is round or elliptical, and has brownish pigment scattered throughout body, a ventral sucker larger than the oral sucker, an elliptical genital sucker lying closely right lateral posteriorly to the ventral sucker, and an O-shaped excretory bladder. Scale bar = 100 µm. (B) *Heterophyopsis continua* metacercaria collected from a yellowfin goby. It is rounded, and has a thick cyst wall (about 6 µm thickness), a ventral sucker larger than the oral sucker, an elliptical genital sucker lying dextroposteriorly to the ventral sucker, and a Y-shaped excretory bladder. Scale bar = 100 µm.



Fig. 11. (A) *Pygidiopsis summa* metacercaria detected from a mullet, *Mugil cephalus*. It is elliptical, and has a pair of eyespots, a genital apparatus lying to the dextroanterior margin of the ventral sucker and an X-shaped excretory bladder. Scale bar = 100 µm. (B) *Stelantchasmus falcatus* metacercaria detected from a mullet. It is elliptical, and has a ventral sucker lying dextrally to median between 2 ceca and a genital atrium lying to the dextrolateral margin of the ventral sucker. Scale bar = 100 µm.

are elliptical, 0.24-0.29 × 0.23-0.28 mm in size, and have a pair of eyespots, genital apparatus crescent formed and lying to the dextroanterior margin of the ventral sucker, and an X-shaped excretory bladder (Fig. 11A). *S. falcatus* metacercariae are elliptical, 0.15-0.20 × 0.13-0.15 mm in size, and have a ventral sucker lying dextrally to median between 2 ceca, a genital atrium lying to the dextrolateral margin of the ventral sucker (Fig. 11B). *S. fuscata* metacercariae are elliptical, 0.19-0.52 × 0.16-0.38 mm in size, and have a very thin and transparent cyst wall and an elliptical gonotyl armed with 12-15 chitinous spines (Fig. 12A). *S. lari* metacercariae are long elliptical, 0.39-0.43 × 0.32-0.35 mm in size, and have a very thin and transparent cyst wall and a gonotyl armed with 60-80 small spines (Fig. 12B). *Acanthotrema felis* metacercariae are elongated elliptical, 0.22-0.25 × 0.15-0.19 mm in size, and have a cyst wall very thin and transparent and a ventrogenital sac which is elliptical and equipped with fork-like sclerites (Fig. 13).



Fig. 12. (A) *Stictodora fuscata* metacercaria detected from a yellowfin goby. It is elliptical, 0.19-0.52 × 0.16-0.38 mm in size, have a very thin and transparent cyst wall and an elliptical gonotyl armed with 12-15 chitinous spines. Scale bar = 100 µm. (B) *Stictodora lari* metacercaria detected from a yellowfin goby. It is elongated and elliptical, 0.39-0.43 × 0.32-0.35 mm in size, have a very thin and transparent cyst wall and a gonotyl armed with 60-80 small spines. Scale bar = 100 µm.

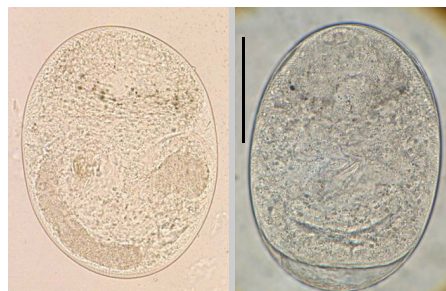


Fig. 13. *Acanthotrema felis* metacercaria detected from a yellowfin goby. It is elongated elliptical and has a very thin and transparent cyst wall and a ventrogenital sac enclosing fork-like sclerites. Scale bar = 100 µm.

Differential keys for metacercariae in brackish water fish

- 1.a. Excretory bladder black and distinctly present 2
 1.b. Excretory bladder vague or absent 5
 2.a. Ventral sucker prominent, genital sucker present 3
 2.b. Ventral sucker smaller, genital sucker absent 4
 3.a. Elliptical, with thin cyst wall and brownish pigment
 *Heterophyes nocens*
 3.b. Rounded, with thick cyst wall *Heterophyopsis continua*
 4.a. Genital apparatus located at anterior margin of ventral sucker, x-shaped excretory bladder *Pygidiopsis summa*
 4.b. Genital atrium with long expulser (seminal vesicle)
 *Stellantchasmus falcatus*
 5.a. Long elliptical, 0.39-0.43 × 0.32-0.35 mm in size, cyst wall very thin and transparent, gonotyl elliptical, armed with 60-80 small spines *Stictodora lari*
 5.b. Elliptical, 0.19-0.52 × 0.16-0.38 mm in size, cyst wall

very thin and transparent, gonotyl elliptical, armed with 12-15 chitinous spines *Stictodora fuscata*

- 5.c. Long elliptical, 0.22-0.25 × 0.15-0.19 mm in size, cyst wall very thin and transparent, ventrogenital sac elliptical, enclosed fork-like sclerites *Acanthotrema felis*

The differential keys for digenetic trematode metacercariae are intended to assist identification of metacercariae recovered in fish from the Republic of Korea. It is not complete to cover all members of trematodes. However, it can be helpful for the identification of FBT metacercariae encountered in fish in the Republic of Korea.

FISH HOSTS

FBT metacercariae encysted in freshwater fish mainly belong to 4 families, Opisthorchiidae, Heterophyidae, Echinostomatidae

Table 3. The fish intermediate hosts of *Clonorchis sinensis* in the Republic of Korea

Family	Genus	Species ^{a)} (= valid name)
Cyprinidae	<i>Abbottina</i>	<i>A. rivularis</i> ¹⁴⁾ , <i>A. springeri</i> ²⁵⁾
	<i>Acanthorhodeus</i>	<i>A. gracilis</i> ²⁵⁾ (= <i>A. chankaensis</i>)
		<i>A. asmussi</i> ¹⁷⁾ (= <i>Acheilognathus asmussi</i>)
	<i>Acheilognathus</i>	<i>A. lanceolata</i> ²⁶⁾ (= <i>Tanakia lanceolata</i>),
		<i>A. signifer</i> ²⁶⁾ , <i>A. rhombeus</i> ²¹⁾ , <i>A. yamatsutae</i> ²⁶⁾
	<i>Aphyocypris</i>	<i>A. chinensis</i> ²⁴⁾
	<i>Carassius</i>	<i>C. auratus</i> ¹⁴⁾
	<i>Coreoleuciscus</i>	<i>C. splendidus</i> ²⁴⁾
	<i>Culter</i>	<i>C. brevicauda</i> ¹⁷⁾ (= <i>Chanodichthys erythropterus</i>)
	<i>Cyprinus</i>	<i>C. carpio</i> ¹⁸⁾
	<i>Erythroculter</i>	<i>E. erythropterus</i> ²¹⁾ (= <i>Chanodichthys erythropterus</i>)
	<i>Gnathopogon</i>	<i>G. strigatus</i> ²⁴⁾
	<i>Hemiculter</i>	<i>H. eigenmanni</i> ²⁴⁾ (= <i>H. leucisculus</i>), <i>H. leucisculus</i> ²⁰⁾
	<i>Hemibarbus</i>	<i>H. labeo</i> ¹⁹⁾ , <i>H. longirostris</i> ¹⁸⁾
	<i>Microphysogobio</i>	<i>M. koreensis</i> ²⁶⁾ , <i>M. yaluensis</i> ²⁴⁾
	<i>Opsarichthys</i>	<i>O. uncirostris amurensis</i> ²⁶⁾ (= <i>O. uncirostris</i>)
	<i>Phoxinus</i>	<i>P. oxycephalus</i> ²⁴⁾
	<i>Pseudogobio</i>	<i>P. esocinus</i> ¹⁷⁾
	<i>Pseudorasbora</i>	<i>P. parva</i> ¹⁴⁾
	<i>Puntungia</i>	<i>P. herzi</i> ¹⁵⁾
	<i>Rhodeus</i>	<i>R. ocellatus</i> ²⁰⁾ (= <i>R. ocellatus ocellatus</i>)
	<i>Saurogobio</i>	<i>S. dabryi</i> ²⁷⁾
	<i>Sarcocheilichthys</i>	<i>S. nigripinnis morii</i> ²⁶⁾ , <i>S. variegatus wakiyae</i> ¹⁸⁾
	<i>Squaliobarbus</i>	<i>S. curriculum</i> ²⁶⁾
	<i>Squalidus</i>	<i>S. japonicus coreanus</i> ¹⁵⁾ , <i>S. gracilis majimae</i> ¹⁶⁾
	<i>Tribolodon</i>	<i>T. hakonensis</i> ¹⁷⁾
	<i>Zacco</i>	<i>Z. platypus</i> ¹⁷⁾ , <i>Z. temminckii</i> ¹⁷⁾
Bagridae	<i>Coreobagrus</i>	<i>C. brevicorpus</i> ²²⁾
Pristigasteridae	<i>Ilisha</i>	<i>I. elongata</i> ²⁶⁾
Osphronemidae	<i>Macropodus</i>	<i>M. chinensis</i> ²³⁾ (= <i>M. opercularis</i>)
Percichthyidae	<i>Coreoperca</i>	<i>C. herzi</i> ²³⁾
	<i>Siniperca</i>	<i>S. scherzeri</i> ²⁹⁾
Osmeridae	<i>Hypomesus</i>	<i>H. olidus</i> ²⁸⁾

^{a)}References which first reported as the second intermediate hosts of *C. sinensis* in Korea.

and Clinostomidae. Nowadays, *C. sinensis* of family Opisthorchiidae is the most important FBT in the Republic of Korea. The metacercariae of *C. sinensis* have been detected in 42 species of freshwater fish (31 genera in 6 families). However, according to the FishBase site in internet [12], *Culter brevicauda* and *Erythroculter erythropterus* are treated as the synonym and then their scientific names were changed into *Chanodichthys erythropterus*. *Hemiculter eigenmanni* is treated as the same species as *H. leucisculus*, and *Acheilognathus lanceolata* is changed into *Tanakia lanceolata*. Therefore, total 40 species of freshwater fish (31 genera in 6 families) act as the second intermediate hosts of *C. sinensis* in the Republic of Korea (Table 3). In China, including Taiwan, total 102 species of freshwater fish of 59 genera in 15 families have been reported as the second intermediate hosts for *C. sinensis* [30].

As members of the Heterophyidae, 3 *Metagonimus* spp. (*M. yokogawai*, *M. takahashii*, and *M. miyatai*) and *Centrocestus armatus* are prevalent in freshwater fish in the Republic of Korea, and their fish intermediate hosts are listed in Table 4. Only 3 fish species (*Plecoglossus altivelis*, *Tribolodon taczanowskii*, and *Lateolabrax japonicus*) have been reported as the second intermediate hosts for *M. yokogawai*, and 4 (*Carassius auratus*, *P. altivelis*, *T. taczanowskii*, and *L. japonicus*) and 2 fish species (*Zacco platypus* and *Z. temminckii*) listed as hosts for *M. takahashii* and *M. miyatai*, respectively. The fish intermediate hosts of *M. yokogawai* are the same species as those of *M. takahashii*, except *C. auratus*. *M. miyatai* revealed a host-specificity at the selection of the second intermediate host. Many fish species have been listed as hosts of unknown *Metagonimus* sp. Studies on differentiation of *Metagonimus*

Table 4. The fish intermediate hosts of heterophyid trematodes in Korea

Trematode species	The second intermediate hosts (References) ^{a)}
<i>Metagonimus yokogawai</i>	<i>Plecoglossus altivelis</i> (Chun, 1960a) ³¹⁾ ; <i>Tribolodon taczanowskii</i> (Choi et al., 1966) ³²⁾ ; <i>Lateolabrax japonicus</i> (Ahn, 1983) ³³⁾
<i>Metagonimus takahashii</i>	<i>Carassius auratus</i> (Chun, 1960b) ³⁴⁾ ; <i>T. taczanowskii</i> (Chai et al., 1991) ³⁵⁾ ; <i>P. altivelis</i> (Rim et al., 1996) ³⁶⁾ ; <i>L. japonicus</i> (Kim et al., 2006) ³⁷⁾
<i>Metagonimus miyatai</i>	<i>Zacco platypus</i> , <i>Zacco temminckii</i> (Saito et al., 1997) ³⁸⁾
<i>Metagonimus</i> spp.	<i>C. auratus</i> , <i>Coreoleuciscus splendidus</i> , <i>Cyprinus carpio</i> , <i>Hemibarbus labeo</i> , <i>Pseudogobio esocinus</i> , <i>Pseudorasbora parva</i> , <i>Puntungia herzi</i> , <i>Z. platypus</i> , <i>Sarcocheilichthys variegatus wakiyae</i> , <i>Squalidus chankaensis tsuchigae</i> (= <i>S. gracilis gracilis</i>), <i>Z. platypus</i> (Lee, 1968) ¹⁹⁾ ; <i>Coreobagrus brevicorpus</i> , <i>Lepomis macrochirus</i> , <i>Macropodus chinensis</i> (= <i>M. opercularis</i>), <i>Acheilognathus rhombeus</i> , <i>Z. temminckii</i> (Joo, 1980) ²²⁾ ; <i>Acheilognathus lanceolata</i> (= <i>Tanakia lanceolata</i>), <i>Coreoperca herzi</i> , <i>Gobiobotia breviparba</i> , <i>Gobiobotia andersonii</i> , <i>Odontobutis macrocephala</i> , <i>Hemibarbus longirostris</i> , <i>Liobagrus platycephala</i> , <i>Pelteobagrus fulvidraco</i> , <i>Pseudopuntungia nigra</i> , <i>Siniperca scherzeri</i> (Kim, 1980) ³⁹⁾ ; <i>Moroco oxycephalus</i> (= <i>Phoxinus oxycephalus</i>) (Kim and Choi, 1981) ⁴⁰⁾ ; <i>Abbottina revularis</i> , <i>A. springeri</i> , <i>Acanthorhodeus gracilis</i> (= <i>A. chankaensis</i>), <i>A. yamatsutae</i> , <i>A. chinensis</i> , <i>Cobitis koreensis</i> (= <i>Iksookimia koreensis</i>), <i>Cobitis taenia</i> , <i>Hemiculter eigenmanni</i> (= <i>H. leucisculus</i>), <i>S. gracilis majimae</i> , <i>G. strigatus</i> , <i>Rhinogobius brunneus</i> , <i>Liobagrus mediadiposalis</i> , <i>M. yaluensis</i> , <i>Rhodeus uyeikii</i> , <i>R. ocellatus ocellatus</i> , <i>S. nigripinnis morii</i> (Rhee et al., 1983) ²⁴⁾ ; <i>Misgurnus anguillicaudatus</i> , <i>Acanthorhodeus asmussi</i> (= <i>Acheilognathus asmussi</i>) (Rhee et al., 84) ²⁵⁾ ; <i>Opsarichthys bidens</i> (Kim et al., 1987) ⁴¹⁾
<i>Centrocestus armatus</i>	<i>A. chinensis</i> , <i>C. auratus</i> , <i>C. splendidus</i> , <i>M. yaluensis</i> , <i>R. uyeikii</i> , <i>P. parva</i> , <i>Z. platypus</i> (Rhee et al., 1983) ²⁴⁾ ; <i>P. fulvidraco</i> , <i>R. ocellatus ocellatus</i> (Rhee et al., 1984) ²⁵⁾ ; <i>Z. temminckii</i> (Hong et al., 1989) ⁴²⁾

^{a)}References that first reported as the second intermediate hosts in Korea.

Table 5. The fish intermediate hosts of echinostomatid trematodes in Korea

Trematode species	The second intermediate hosts (References) ^{a)}
<i>Echinostoma hortense</i>	<i>Misgurnus anguillicaudatus</i> (Chai et al., 1985) ⁴³⁾ ; <i>Moroco oxycephalus</i> (= <i>Phoxinus oxycephalus</i>) (Ryang et al., 1985) ⁴⁴⁾ ; <i>Odontobutis obscura interrupta</i> (= <i>O. interrupta</i>) (Ahn and Ryang, 1986) ⁴⁵⁾ ; <i>Squalidus coreanus</i> (= <i>S. japonicus coreanus</i>) (Lee et al., 1988) ⁴⁶⁾ ; <i>Rhinogobius brunneus</i> , <i>Acanthorhodeus macropterus</i> (= <i>Acheilognathus macropterus</i>) (Rim et al., 1996) ⁴⁷⁾ ; <i>Acanthogobius flavimanus</i> (Sohn et al., 2009) ⁴⁸⁾
<i>Echinostoma cinetorchis</i>	<i>Misgurnus anguillicaudatus</i> (Seo et al., 1984) ⁴⁹⁾
<i>Echinochasmus japonicus</i>	<i>Pseudorasbora parva</i> , <i>Hypomesus olidus</i> (Rhee et al., 1983) ²⁴⁾ ; <i>Squalidus japonicus coreanus</i> , <i>Gnathopogon strigatus</i> , <i>Squalidus gracilis majimae</i> , <i>Pungtungia herzi</i> , <i>Zacco platypus</i> , <i>Abbottina springeri</i> , <i>Acheilognathus lanceolata</i> (= <i>Tanakia lanceolata</i>), <i>Aphyocypris chinensis</i> , <i>Carassius auratus</i> , <i>Hemibarbus longirostris</i> , <i>Moroco oxycephalus</i> (= <i>Phoxinus oxycephalus</i>), <i>Pseudogobio esocinus</i> , <i>Rhodeus uyeikii</i> , <i>Acanthorhodeus gracilis</i> (= <i>A. chankaensis</i>), <i>Pelteobagrus fulvidraco</i> (Rhee et al., 1984) ²⁵⁾

^{a)}References that first reported as the second intermediate hosts in Korea.

Table 6. The second intermediate hosts of heterophyid trematodes in Korea

Trematode species	The second intermediate hosts (References) ^{a)}
<i>Heterophyes nocens</i>	<i>Mugil cephalus</i> (Seo et al., 1980) ⁵¹⁾ ; <i>Acanthogobius flavimanus</i> (Seo et al., 1981) ⁵²⁾ ; <i>Boleophthalmus pectinirostris</i> , <i>Scartelaos</i> sp. (Sohn et al., 2005) ⁵³⁾ ; <i>Chelon haematocheilus</i> (Kim et al. 2006) ⁵⁴⁾
<i>Heterophyopsis continua</i>	<i>Laterolabrax japonicus</i> (Chun, 1960c) ⁵⁵⁾ ; <i>A. flavimanus</i> (Seo et al., 1984) ⁵⁶⁾ ; <i>Clupanodon punctatus</i> (Sohn et al., 1994a) ⁵⁷⁾ ; <i>Plecoglossus altivelis</i> (Cho and Kim, 1985) ⁵⁸⁾ ; <i>Conger myriaster</i> (Kim et al., 1996) ⁵⁹⁾ ; <i>B. pectinirostris</i> , <i>Scartelaos</i> sp. (Sohn et al., 2005) ⁵³⁾
<i>Pygidiopsis summa</i>	<i>M. cephalus</i> (Chun, 1963) ⁶⁰⁾ ; <i>A. flavimanus</i> (Seo et al., 1981) ⁶¹⁾ ; <i>C. haematocheilus</i> (Kim et al. 2006) ⁵⁴⁾
<i>Stellantchasmus falcatus</i>	<i>M. cephalus</i> (Chai and Sohn, 1988) ⁶²⁾
<i>Stictodora fuscata</i>	<i>A. flavimanus</i> (Sohn et al., 1994b) ⁶³⁾ ; <i>B. pectinirostris</i> (Sohn et al., 2005) ⁵³⁾
<i>Stictodora lari</i>	<i>A. flavimanus</i> (Chai et al., 1989) ⁶⁴⁾
<i>Acanthotrema felis</i>	<i>A. flavimanus</i> (Sohn et al., 2003) ⁶⁵⁾

^{a)}References that first reported as the second intermediate hosts in Korea.

sp. metacercariae are needed.

There are 3 species of echinostomatid metacercariae, i.e., *Echinostoma hortense*, *Echinostoma cinetorchis*, and *Echinostomus japonicus*, found from freshwater fish in the Republic of Korea (Table 5). *Clinostomum complanatum* metacercariae were found in 12 species of freshwater fish, *Acheilognathus koreensis*, *Acheilognathus rhombea* (= *A. rhombeus*), *Acheilognathus yamatsutae*, *Carassius auratus*, *Cobitis sinensis*, *Microphysogobio yaluensis*, *Pseudorasbora parva*, *Pungtungia herzi*, *Rhodeus uyekii*, *Squalidus chankaensis tsuchigae* (= *S. gracilis gracilis*), *Squalidus gracilis maejimai*, and *Zacco temminkii*, as reported by Chung et al. [49].

FBT metacercariae detected in brackish water fish are mainly those of the Heterophyidae, including *Heterophyes nocens*, *Heterophyopsis continua*, *Pygidiopsis summa*, *Stellantchasmus falcatus*, *Stictodora fuscata*, *S. lari*, and *Acanthotrema felis*. Their second intermediate hosts are designated in Table 6.

CONCLUSION

The detection and identification methods, differential keys, the second intermediate hosts, and morphological characteristics of fish-borne zoonotic trematode metacercariae in the Republic of Korea were briefly reviewed. The items treated in this paper will be useful to understand the epidemiology of fish-borne trematodiasis in the Republic of Korea, and to provide assistance at the laboratory bench level to those who work on metacercariae in fish hosts.

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